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| 10/564,512 | 05/12/2006 | Jean-Paul Di Rago | 284469US0XPCT | 3450 |
| 22850 | 7590 | 06/21/2010 | EXAMINER | |
| OBLON, SPIVAK, MCCLELLAND MAIER & NEUSTADT, L.L.P. 1940 DUKE STREET ALEXANDRIA, VA 22314 | | | JOIKE, MICHELE K | |
| | | | ART UNIT | PAPER NUMBER |
| | | | 1636 | |
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Please find below and/or attached an Office communication concerning this application or proceeding.

The time period for reply, if any, is set in the attached communication.

Notice of the Office communication was sent electronically on above-indicated "Notification Date" to the following e-mail address(es):

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|------------------------------|--------------------------------------|---------------------------------------|--|
| Office Action Summary | Application No. 10/564,512 | Applicant(s) DI RAGO ET AL. | |
| | Examiner Michele K. Joiike | Art Unit 1636 | |

-- The MAILING DATE of this communication appears on the cover sheet with the correspondence address --

Period for Reply

A SHORTENED STATUTORY PERIOD FOR REPLY IS SET TO EXPIRE 3 MONTH(S) OR THIRTY (30) DAYS, WHICHEVER IS LONGER, FROM THE MAILING DATE OF THIS COMMUNICATION.

- Extensions of time may be available under the provisions of 37 CFR 1.136(a). In no event, however, may a reply be timely filed after SIX (6) MONTHS from the mailing date of this communication.
- If NO period for reply is specified above, the maximum statutory period will apply and will expire SIX (6) MONTHS from the mailing date of this communication.
- Failure to reply within the set or extended period for reply will, by statute, cause the application to become ABANDONED (35 U.S.C. § 133). Any reply received by the Office later than three months after the mailing date of this communication, even if timely filed, may reduce any earned patent term adjustment. See 37 CFR 1.704(b).

Status

- 1) ☒ Responsive to communication(s) filed on 10 March 2010.
- 2a) ☐ This action is **FINAL**. 2b) ☒ This action is non-final.
- 3) ☐ Since this application is in condition for allowance except for formal matters, prosecution as to the merits is closed in accordance with the practice under *Ex parte Quayle*, 1935 C.D. 11, 453 O.G. 213.

Disposition of Claims

- 4) ☒ Claim(s) 1-14 and 21-24 is/are pending in the application.
- 4a) Of the above claim(s) _____ is/are withdrawn from consideration.
- 5) ☐ Claim(s) _____ is/are allowed.
- 6) ☒ Claim(s) 1-14 and 21-23 is/are rejected.
- 7) ☒ Claim(s) 24 is/are objected to.
- 8) ☐ Claim(s) _____ are subject to restriction and/or election requirement.

Application Papers

- 9) ☐ The specification is objected to by the Examiner.
- 10) ☐ The drawing(s) filed on _____ is/are: a) ☐ accepted or b) ☐ objected to by the Examiner.
Applicant may not request that any objection to the drawing(s) be held in abeyance. See 37 CFR 1.85(a).
Replacement drawing sheet(s) including the correction is required if the drawing(s) is objected to. See 37 CFR 1.121(d).
- 11) ☐ The oath or declaration is objected to by the Examiner. Note the attached Office Action or form PTO-152.

Priority under 35 U.S.C. § 119

- 12) ☐ Acknowledgment is made of a claim for foreign priority under 35 U.S.C. § 119(a)-(d) or (f).
- a) ☐ All b) ☐ Some * c) ☐ None of:
1. ☐ Certified copies of the priority documents have been received.
 2. ☐ Certified copies of the priority documents have been received in Application No. _____.
 3. ☐ Copies of the certified copies of the priority documents have been received in this National Stage application from the International Bureau (PCT Rule 17.2(a)).

* See the attached detailed Office action for a list of the certified copies not received.

Attachment(s)

- | | |
|--|---|
| 1) <input checked="" type="checkbox"/> Notice of References Cited (PTO-892) | 4) <input type="checkbox"/> Interview Summary (PTO-413) |
| 2) <input type="checkbox"/> Notice of Draftsperson's Patent Drawing Review (PTO-948) | Paper No(s)/Mail Date. _____ |
| 3) <input type="checkbox"/> Information Disclosure Statement(s) (PTO/SB/08) | 5) <input type="checkbox"/> Notice of Informal Patent Application |
| Paper No(s)/Mail Date _____ | 6) <input type="checkbox"/> Other: _____ |

DETAILED ACTION

Receipt is acknowledged of a reply to the previous Office Action, filed March 10, 2010. Claims 1-14 and 21-24 are pending and under consideration in the instant application.

Any rejection of record in the previous Office Action, mailed September 10, 2009 that is not addressed in this action has been withdrawn. Because this Office Action introduces new rejections other than those set forth in the previous Office Action, and are not necessitated by amendment, this Office Action is Non-Final.

Response to Arguments

Applicant's arguments regarding Bonnefoy et al , see page 10, and Kasiha et al, see page 13, filed March 10, 2010, with respect to the rejection(s) of claim(s) 1, 2, 5, 7, 8, 11 and 12 under 35 USC 103(a) have been fully considered and are persuasive. Therefore, the rejection has been withdrawn. However, upon further consideration, a new ground(s) of rejection is made.

Claim Rejections - 35 USC § 102

The following is a quotation of the appropriate paragraphs of 35 U.S.C. 102 that form the basis for the rejections under this section made in this Office action:

A person shall be entitled to a patent unless –

(b) the invention was patented or described in a printed publication in this or a foreign country or in public use or on sale in this country, more than one year prior to the date of application for patent in the United States.

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Claims 1, 2, 5, 8-10, 21, 22 and 23 are rejected under 35 U.S.C. 102(b) as being anticipated by Bonnefoy et al.

Bonnefoy et al (Mol. Gen. Genet. 262: 1036-1046, 2000, see entire paper, hereinafter Bonnefoy 2000) teach transformation of *S. cerevisiae* mitochondria. The mitochondria are transformed with a mitochondria transcription vector carrying an ARG8 reporter gene and *cox2* (1-91). ARG8 is an auxotrophic mutant that can be expressed in mitochondria. The mitochondria were also co-transformed with a LEU2 plasmid to select nuclear transformants. The mitochondria are transformed by microprojectile bombardment. Cells that survive are selected for. Strains that can be used in the transformation are rho- (large deletions of mtDNA) or rho°(lacking mtDNA). Bonnefoy et al teach using an auxotrophic or drug resistance marker, or a deletion in a region of interest that will allow respiring recombinants to grow, to confirm transformation. Yeast mitochondria are isolated and RNA is extracted and purified. Absent evidence to the contrary, the DNA encoding the RNA was under control of a promoter and terminator that are functional in yeast mitochondria, since the RNA was successfully produced in yeast mitochondria.

Claim Rejections - 35 USC § 103

The following is a quotation of 35 U.S.C. 103(a) which forms the basis for all obviousness rejections set forth in this Office action:

(a) A patent may not be obtained though the invention is not identically disclosed or described as set forth in section 102 of this title, if the differences between the subject matter sought to be patented and the prior art are such that the subject matter as a whole would have been obvious at the time the invention was made to a person having ordinary skill in the art to which said subject matter pertains. Patentability shall not be negated by the manner in which the invention was made.

This application currently names joint inventors. In considering patentability of the claims under 35 U.S.C. 103(a), the examiner presumes that the subject matter of the various claims was commonly owned at the time any inventions covered therein were made absent any evidence to the contrary. Applicant is advised of the obligation under 37 CFR 1.56 to point out the inventor and invention dates of each claim that was not commonly owned at the time a later invention was made in order for the examiner to consider the applicability of 35 U.S.C. 103(c) and potential 35 U.S.C. 102(e), (f) or (g) prior art under 35 U.S.C. 103(a).

Claims 1, 2, 5, 7, 8, 11 and 21-23 are rejected under 35 U.S.C. 103(a) as being unpatentable over Bonnefoy et al (hereinafter Bonnefoy 2001) in view of US 2009/0098653.

Bonnefoy 2001 (IDS reference AX, especially pp. 98-101, 104-105 and 109) teach transformation of *S. cerevisiae* mitochondria. The mitochondria are transformed with a vector carrying an ARG8 reporter gene. Since the vector is transformed and expressed in mitochondria, it is a mitochondrial transcription vector. ARG8 is an auxotrophic mutant that can be expressed in mitochondria. The mitochondria are transformed by microprojectile bombardment. Cells that survive are selected for. Strains that can be used in the transformation are rho- (large deletions of mtDNA) or rho°(lacking mtDNA). A tester strain can also be used, rho+, mit-. The rho+ tester strain can be mated to a rho- strain. After mating, diploids will be produced when grown on a non-fermentable medium, and they teach that the cells can be mated twice. Also, step c₀ implies that the crossing does not have to be repeated if the colonies are

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identified as being mitochondrial transformants. Bonnefoy et al teach using an auxotrophic or drug resistance marker, or a deletion in a region of interest that will allow respiring recombinants to grow, to confirm transformation. They also teach transforming rho^o strains with a plasmid containing an origin of replication that allows the plasmid to replicate. Absent evidence to the contrary, the DNA encoding the protein was under control of a promoter and terminator that are functional in yeast mitochondria, since the protein was successfully produced in yeast mitochondria. However, they do not teach producing a heterologous RNA.

US 2009/0098653 (especially Ex. 13-15) teach transforming mitochondria lacking mitochondria DNA with a plasmid comprising a gene which is expressed to produce RNA. The mitochondria were isolated, and then the RNA was isolated from the mitochondria.

The ordinary skilled artisan, desiring to produce RNA in yeast mitochondria, would have been motivated to combine the teachings of Bonnefoy 2001 teaching transformation of *S. cerevisiae* mitochondria with the teachings of US 2009/0098653 teaching transforming mitochondria with a plasmid comprising a gene which is expressed to produce RNA because Bonnefoy 2001 state that genetic manipulation of *S. cerevisiae* mitochondria are amenable to in vivo experimental analysis and should provide a useful model for other systems. It would have been obvious to one of ordinary skill in the art to use mitochondria to produce RNA because US 2009/0098653 teach that transforming yeast mitochondria with exogenous DNA allows for studying of transcription, and production of mRNA in the mitochondria. The methods also teach a

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means for RT-PCR, which allows for production of RNA in mitochondria. Given the teachings of the prior art and the level of the ordinary skilled artisan at the time of the applicant's invention, it must be considered, absent evidence to the contrary, that said skilled artisan would have had a reasonable expectation of success in practicing the claimed invention.

Claim 3 is rejected under 35 U.S.C. 103(a) as being unpatentable over Bonnefoy 2001 and US 2009/0098653 as applied to claims 1, 2, 5, 7, 8, 11 and 21-23 above, and further in view of Dziembowski et al.

Bonnefoy 2001 and US 2009/0098653 teach all of the limitations as described above. However, they do not teach using Δ SUV3 or Δ DSS1 strains.

Dziembowski et al (J. Biol. Chem. 278(3): 1603-1611, 2003, especially p. 1603) teach using Δ SUV3 or Δ DSS1 strains.

The ordinary skilled artisan, desiring to use Δ SUV3 or Δ DSS1 strains in RNA production in yeast mitochondria, would have been motivated to combine the teachings of Bonnefoy 2001 teaching transformation of *S. cerevisiae* mitochondria with the teachings of US 2009/0098653 teaching transforming mitochondria with a plasmid comprising a gene which is expressed to produce RNA, with Dziembowski et al teaching using Δ SUV3 or Δ DSS1 strains because Dziembowski et al state that inactivation of SUV3 or DSS1 results in respiratory incompetence and eventual loss of the mitochondrial genome. It would have been obvious to one of ordinary skill in the art to use Δ SUV3 or Δ DSS1 strains because Dziembowski et al teach that inactivation of

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SUV3 or DSS1 leads to strong inhibition of mitochondrial translation. Given the teachings of the prior art and the level of the ordinary skilled artisan at the time of the applicant's invention, it must be considered, absent evidence to the contrary, that said skilled artisan would have had a reasonable expectation of success in practicing the claimed invention.

Claim 4 is rejected under 35 U.S.C. 103(a) as being unpatentable over Bonnefoy 2001 and US 2009/0098653 as applied to claims 1, 2, 5, 7, 8, 11 and 21-23 above, and further in view of Komiya et al and Hwang et al.

Bonnefoy 2001 and US 2009/0098653 teach all of the limitations as described above. However, they do not teach cells having a chromosomal copy of a gene encoding an RNAP, or a mitochondrial targeting signal.

Hwang et al (J. of Virology 74(9): 4074-4084, 2000, especially p. 4075) teach a viral RNAP integrated into the genome of Pichia. However, they do not teach cells having a mitochondrial targeting signal.

Komiya et al (J. Biol. Chem. 269(49): 30893-30897, 1994, especially 30896) teach using a mitochondrial targeting signal for cytosolic import.

The ordinary skilled artisan, desiring to use a cell having a chromosomal copy of a gene encoding an RNAP and a mitochondrial targeting signal, would have been motivated to combine the teachings of Bonnefoy 2001 teaching transformation of *S. cerevisiae* mitochondria with the teachings of US 2009/0098653 teaching transforming mitochondria with a plasmid comprising a gene which is expressed to produce RNA,

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with the teachings of Hwang et al and Komiya et al because Hwang et al state that using this expression system allowed for sufficient amounts of the polymerase, and was easily expressed, at a low cost. It would have been obvious to one of ordinary skill in the art to use a mitochondrial targeting signal because Komiya et al teach that mitochondrial targeting signals are important for the importation of proteins into the mitochondria. Given the teachings of the prior art and the level of the ordinary skilled artisan at the time of the applicant's invention, it must be considered, absent evidence to the contrary, that said skilled artisan would have had a reasonable expectation of success in practicing the claimed invention.

Claim 6 is rejected under 35 U.S.C. 103(a) as being unpatentable over Bonnefoy 2001 and US 2009/0098653 as applied to claims 1, 2, 5, 7, 8, 11 and 21-23 above, and further in view of Anziano et al.

Bonnefoy 2001 and US 2009/0098653 teach all of the limitations as described above. However, they do not teach that the reporter gene is gene encoding a protein from the yeast respiratory chain.

Anziano et al (IDS ref. AW, especially p. 5396) teach use of the COXII gene as a reporter.

The ordinary skilled artisan, desiring to use a reporter gene that is a gene encoding a protein from the yeast respiratory chain, would have been motivated to combine the teachings of Bonnefoy 2001 teaching transformation of *S. cerevisiae* mitochondria with the teachings of US 2009/0098653 teaching transforming

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mitochondria with a plasmid comprising a gene which is expressed to produce RNA, with Anziano al teaching transformation of the COXII gene into yeast, because Anziano et al state that the COXII gene is a convenient selectable marker for primary mitochondrial transformants. It would have been obvious to one of ordinary skill in the art to use the COXII gene as a reporter because Anziano et al teach that now make it possible, to express proteins in mitochondria without having to select directly for them, or to resort to engineering their expression and subsequent import into mitochondria in the nucleus-cytoplasm. Given the teachings of the prior art and the level of the ordinary skilled artisan at the time of the applicant's invention, it must be considered, absent evidence to the contrary, that said skilled artisan would have had a reasonable expectation of success in practicing the claimed invention.

Claims 9, 10 and 12 are rejected under 35 U.S.C. 103(a) as being unpatentable over Bonnefoy 2001 and US 2009/0098653 as applied to claims 1, 2, 5, 7, 8, 11 and 21-23 above, and further in view of Fincham.

Bonnefoy 2001 and US 2009/0098653 teach all of the limitations as described above. However, they do not teach co-transformation.

Fincham (Micro. Rev. 53(1): 148-170, 1989, especially p. 151) teaches co-transformation in yeast. It also teaches various markers, including nuclear markers that can be used for selection.

The ordinary skilled artisan, desiring to co-transform plasmids, would have been motivated to combine the teachings of Bonnefoy 2001 teaching transformation of S.

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cerevisiae mitochondria with the teachings of US 2009/0098653 teaching transforming mitochondria with a plasmid comprising a gene which is expressed to produce RNA, with Fincham teaching co-transformation in yeast, because Fincham states that co-transformation is useful for when a gene cannot easily be directly selected. It would have been obvious to one of ordinary skill in the art to use co-transformation because there is a high probability that if the cell will take upon plasmid, it will also take up the second plasmid. Given the teachings of the prior art and the level of the ordinary skilled artisan at the time of the applicant's invention, it must be considered, absent evidence to the contrary, that said skilled artisan would have had a reasonable expectation of success in practicing the claimed invention.

Claim 13 is rejected under 35 U.S.C. 103(a) as being unpatentable over Bonnefoy 2001 and US 2009/0098653 as applied to claims 1, 2, 5, 7, 8, 11 and 21-23 above, and further in view of Kim et al.

Bonnefoy 2001 and US 2009/0098653 teach all of the limitations as described above.

Kim et al (Cancer Res. 57: 3115-3120, 1997, especially p. 3116) teach lysing cells and centrifuging twice at 750 x g to isolate mitochondria.

The claim would have been obvious because centrifuging twice instead of once is a simple adjustment that was known in the art and would have yielded predictable results to one of skill in the art at the time of the invention.

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Claim 14 is rejected under 35 U.S.C. 103(a) as being unpatentable over Bonnefoy 2001 and US 2009/0098653 as applied to claims 1, 2, 5, 7, 8, 11 and 21-23 above, and further in view of Dziembowski et al and di Rago et al.

Bonnefoy 2001 and US 2009/0098653 teach all of the limitations as described above. However, they do not teach eliminating the contaminating nucleic acids in the presence of a divalent ion-chelating agent and a second buffer comprising RNase.

Dziembowski et al teach all of the limitations as described above. They also teach lysing mitochondria with a detergent, Triton X-100 and EDTA.

Di Rago et al (J. Biol. Chem. 263(25): 12564-12570, 1988, especially p. 12565) teach isolating mitochondrial RNA using buffers containing EDTA pH 7.4, and DNase. They teach using DNase, but not RNase.

The claim would have been obvious because the buffers used in the references above were well known in the art for lysing of cells and organelles, and isolating RNA. The claim would have been obvious because the substitution of one known element (RNase) for another (DNase) would have yielded predictable results to one of skill in the art at the time of the invention. RNase is well known to degrade RNA.

Claim 3 is rejected under 35 U.S.C. 103(a) as being unpatentable over Bonnefoy 2000 as applied to claims 1, 2, 5, 8-10, 21, 22 and 23 above, and further in view of Dziembowski et al.

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Bonnefoy 2000 teach all of the limitations as described above. However, they do not teach using Δ SUV3 or Δ DSS1 strains.

Dziembowski et al (J. Biol. Chem. 278(3): 1603-1611, 2003, especially p. 1603) teach using Δ SUV3 or Δ DSS1 strains.

The ordinary skilled artisan, desiring to use Δ SUV3 or Δ DSS1 strains in RNA production in yeast mitochondria, would have been motivated to combine the teachings of Bonnefoy 2000 teaching transformation of *S. cerevisiae* mitochondria, with Dziembowski et al teaching using Δ SUV3 or Δ DSS1 strains because Dziembowski et al state that inactivation of SUV3 or DSS1 results in respiratory incompetence and eventual loss of the mitochondrial genome. It would have been obvious to one of ordinary skill in the art to use Δ SUV3 or Δ DSS1 strains because Dziembowski et al teach that inactivation of SUV3 or DSS1 leads to strong inhibition of mitochondrial translation. Given the teachings of the prior art and the level of the ordinary skilled artisan at the time of the applicant's invention, it must be considered, absent evidence to the contrary, that said skilled artisan would have had a reasonable expectation of success in practicing the claimed invention.

Claim 4 is rejected under 35 U.S.C. 103(a) as being unpatentable over Bonnefoy 2000 as applied to claims 1, 2, 5, 8-10, 21, 22 and 23 above, and further in view of Komiya et al and Hwang et al.

Bonnefoy 2000 teach all of the limitations as described above. However, they do not teach cells having a chromosomal copy of a gene encoding an RNAP, or a mitochondrial targeting signal.

Hwang et al (J. of Virology 74(9): 4074-4084, 2000, especially p. 4075) teach a viral RNAP integrated into the genome of Pichia. However, they do not teach cells having a mitochondrial targeting signal.

Komiya et al (J. Biol. Chem. 269(49): 30893-30897, 1994, especially 30896) teach using a mitochondrial targeting signal for cytosolic import.

The ordinary skilled artisan, desiring to use a cell having a chromosomal copy of a gene encoding an RNAP and a mitochondrial targeting signal, would have been motivated to combine the teachings of Bonnefoy 2000 teaching transformation of *S. cerevisiae* mitochondria with the teachings of Hwang et al and Komiya et al because Hwang et al state that using this expression system allowed for sufficient amounts of the polymerase, and was easily expressed, at a low cost. It would have been obvious to one of ordinary skill in the art to use a mitochondrial targeting signal because Komiya et al teach that mitochondrial targeting signals are important for the importation of proteins into the mitochondria. Given the teachings of the prior art and the level of the ordinary skilled artisan at the time of the applicant's invention, it must be considered, absent evidence to the contrary, that said skilled artisan would have had a reasonable expectation of success in practicing the claimed invention.

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Claim 6 is rejected under 35 U.S.C. 103(a) as being unpatentable over Bonnefoy 2000 as applied to claims 1, 2, 5, 8-10, 21, 22 and 23 above, and further in view of Anziano et al.

Bonnefoy 2000 teach all of the limitations as described above. While Bonnefoy 2000 teach use of the cox2 gene, they do not use it as a reporter.

Anziano et al (IDS ref. AW, especially p. 5396) teach use of the COXII gene as a reporter.

The ordinary skilled artisan, desiring to use a reporter gene that is a gene encoding a protein from the yeast respiratory chain, would have been motivated to combine the teachings of Bonnefoy 2000 teaching transformation of *S. cerevisiae* mitochondria with Anziano et al teaching transformation of the COXII gene into yeast, because Anziano et al state that the COXII gene is a convenient selectable marker for primary mitochondrial transformants. It would have been obvious to one of ordinary skill in the art to use the COXII gene as a reporter because Anziano et al teach that now make it possible, to express proteins in mitochondria without having to select directly for them, or to resort to engineering their expression and subsequent import into mitochondria in the nucleus-cytoplasm. Given the teachings of the prior art and the level of the ordinary skilled artisan at the time of the applicant's invention, it must be considered, absent evidence to the contrary, that said skilled artisan would have had a reasonable expectation of success in practicing the claimed invention.

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Claim 13 is rejected under 35 U.S.C. 103(a) as being unpatentable over Bonnefoy 2000 as applied to claims 1, 2, 5, 8-10, 21, 22 and 23 above, and further in view of Kim et al.

Bonnefoy 2000 teach all of the limitations as described above.

Kim et al (Cancer Res. 57: 3115-3120, 1997, especially p. 3116) teach lysing cells and centrifuging twice at 750 x g to isolate mitochondria.

The claim would have been obvious because centrifuging twice instead of once is a simple adjustment that was known in the art and would have yielded predictable results to one of skill in the art at the time of the invention.

Claim 14 is rejected under 35 U.S.C. 103(a) as being unpatentable over Bonnefoy 2000 as applied to claims 1, 2, 5, 8-10, 21, 22 and 23 above, and further in view of Dziembowski et al and di Rago et al.

Bonnefoy 2000 teach all of the limitations as described above. However, they do not teach eliminating the contaminating nucleic acids in the presence of a divalent ion-chelating agent and a second buffer comprising RNase.

Dziembowski et al teach all of the limitations as described above. They also teach lysing mitochondria with a detergent, Triton X-100 and EDTA.

Di Rago et al (J. Biol. Chem. 263(25): 12564-12570, 1988, especially p. 12565) teach isolating mitochondrial RNA using buffers containing EDTA pH 7.4, and DNase. They teach using DNase, but not RNase.

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The claim would have been obvious because the buffers used in the references above were well known in the art for lysing of cells and organelles, and isolating RNA. The claim would have been obvious because the substitution of one known element (RNase) for another (DNase) would have yielded predictable results to one of skill in the art at the time of the invention. RNase is well known to degrade RNA.

Claims 1, 2, 5, 8-11 and 21-23 are rejected under 35 U.S.C. 103(a) as being unpatentable over Bonnefoy 2001 in view of Bonnefoy 2000.

Bonnefoy 2001 (IDS reference AX, especially pp. 98-101, 104-105 and 109) teach transformation of *S. cerevisiae* mitochondria. The mitochondria are transformed with a vector carrying an ARG8 reporter gene. Since the vector is transformed and expressed in mitochondria, it is a mitochondrial transcription vector. ARG8 is an auxotrophic mutant that can be expressed in mitochondria. The mitochondria are transformed by microprojectile bombardment. Cells that survive are selected for. Strains that can be used in the transformation are rho- (large deletions of mtDNA) or rho°(lacking mtDNA). A tester strain can also be used, rho+, mit-. The rho+ tester strain can be mated to a rho- strain. After mating, diploids will be produced when grown on a non-fermentable medium, and they teach that the cells can be mated twice. Also, step c₀ implies that the crossing does not have to be repeated if the colonies are identified as being mitochondrial transformants. Bonnefoy et al teach using an auxotrophic or drug resistance marker, or a deletion in a region of interest that will allow respiring recombinants to grow, to confirm transformation. They also teach

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transforming rho^o strains with a plasmid containing an origin of replication that allows the plasmid to replicate. However, they do not teach co-transformation, or production and isolation of RNA.

Bonnefoy 2000 (Mol. Gen. Genet. 262: 1036-1046, 2000, see entire paper) teach transformation of *S. cerevisiae* mitochondria. The mitochondria are transformed with a mitochondria transcription vector carrying an ARG8 reporter gene and cox2 (1-91). ARG8 is an auxotrophic mutant that can be expressed in mitochondria. The mitochondria were also co-transformed with a LEU2 plasmid to select nuclear transformants. The mitochondria are transformed by microprojectile bombardment. Cells that survive are selected for. Strains that can be used in the transformation are rho⁻ (large deletions of mtDNA) or rho^o (lacking mtDNA). Bonnefoy et al teach using an auxotrophic or drug resistance marker, or a deletion in a region of interest that will allow respiring recombinants to grow, to confirm transformation. Yeast mitochondria are isolated and RNA is extracted and purified.

The ordinary skilled artisan, desiring to produce RNA in yeast mitochondria, would have been motivated to combine the teachings of Bonnefoy 2001 teaching transformation of *S. cerevisiae* mitochondria with the teachings of Bonnefoy 2000 teaching transforming mitochondria with a plasmid comprising a gene which is expressed to produce RNA because Bonnefoy 2001 state that genetic manipulation of *S. cerevisiae* mitochondria are amenable to in vivo experimental analysis and should provide a useful model for other systems. It would have been obvious to one of ordinary skill in the art to use mitochondria to produce RNA because Bonnefoy 2000

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teach that translation initiation in mitochondrial systems is poorly understood, and that examination of mRNA structure and genetic analysis makes it possible to ascertain features of translation initiation in fungal mitochondria. Given the teachings of the prior art and the level of the ordinary skilled artisan at the time of the applicant's invention, it must be considered, absent evidence to the contrary, that said skilled artisan would have had a reasonable expectation of success in practicing the claimed invention.

Allowable Subject Matter

Claim 24 is objected to as being dependent upon a rejected base claim, but would be allowable if rewritten in independent form including all of the limitations of the base claim and any intervening claims.

Any inquiry concerning this communication or earlier communications from the examiner should be directed to Michele K. Joike whose telephone number is (571)272-5915. The examiner can normally be reached on M-F, 10:00-6:30.

If attempts to reach the examiner by telephone are unsuccessful, the examiner's supervisor, Christopher Low can be reached on (571)272-0951. The fax phone number for the organization where this application or proceeding is assigned is 571-273-8300.

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Information regarding the status of an application may be obtained from the Patent Application Information Retrieval (PAIR) system. Status information for published applications may be obtained from either Private PAIR or Public PAIR. Status information for unpublished applications is available through Private PAIR only. For more information about the PAIR system, see <http://pair-direct.uspto.gov>. Should you have questions on access to the Private PAIR system, contact the Electronic Business Center (EBC) at 866-217-9197 (toll-free). If you would like assistance from a USPTO Customer Service Representative or access to the automated information system, call 800-786-9199 (IN USA OR CANADA) or 571-272-1000.

/Michele K. Joike/
Primary Examiner, Art Unit 1636

Michele K. Joike
Primary Examiner
Art Unit 1636